Chlorolab 2+

Liquid-phase oxygen electrode system for advanced photosynthesis & respiration studies





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- > PC operated USB Oxylab+ electrode control unit
- > DW2/2 advanced electrode chamber with 4 optical ports and integral S1 oxygen electrode disc
- > LED1/W white LED light source (up to 2,000 µmols m⁻² s⁻¹) with automated control via user-defined PFD light tables
- > Suitable for liquid-phase samples between 0.2 2.5ml (min. 1.5ml if illuminated) with 0 100% oxygen concentration
- > 24 bit high resolution measurement of oxygen signals
- > Integral systems for measurement of pH & other ionselective electrode (ISE) signals with 16 bit resolution
- > Onboard LCD readings of oxygen, auxiliary & ISE signals
- > 2 channel capability via purchase of additional systems
- > Quantitherm PAR/Temp sensor for light source calibration
- OxyTrace+ Windows® software for data acquisition, hardware control & data analysis
- > Real time 0 4.5v analogue output of oxygen signal



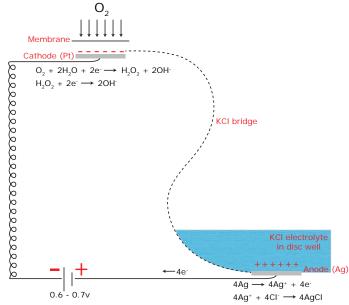
control unit. As part of a complete system, Oxylab+ provides a convenient yet powerful tool for measurements of oxygen evolution or uptake across a broad range of liquid-phase samples from chloroplast extractions to mitochondrial suspensions with oxygen concentrations up to 100%.

Oxylab+ offers unrivalled price vs. performance combining simplicity of operation with an enviable feature set. The outstanding flexibility ensures Oxylab+ is equally useful in both a teaching and research capacity. 24 bit resolution allows detection of minute changes in oxygen tension without needing to apply instrument gain. This results in beautiful, noise free traces even when zoomed close in on areas of interest. Integral electronics provide control over an LED light source with automatic intensity changes handled by user-defined PFD light tables in software.

The system allows realtime graphing of signals from auxiliary inputs and ion-selective electrodes providing scope for comprehensive analysis of oxygen activity simultaneously with signals such as pH, TPP+, calcium, potassium and hydrogen ions. Signals from all inputs are additionally displayed on an LCD screen mounted within the front panel of the Oxylab+ control unit.

Up to 2 individual Oxylab+ control units may be linked to a single PC and operated simultaneously from OxyTrace+ software providing a powerful, multi-channel system.





Oxygen electrode disc

Since its original design in the early 1970's by Tom Delieu and David Walker, the S1 Clark Type Oxygen Electrode disc remains largely unchanged - a true testament to the quality and reliability of the sensor. The S1 consists of a platinum cathode and silver anode set into an epoxy resin disc and is prepared for use by trapping a layer of 50% saturated KCl solution beneath an oxygen permeable PTFE membrane. A paper spacer placed beneath the membrane acts as a wick to provide a uniform layer of electrolyte between anode and cathode.

When a small voltage is applied across these electrodes (with the platinum negative with respect to the silver), the current which flows is at first negligible and the platinum becomes polarised (i.e. it adopts the externally applied potential). As this potential is increased to 700 mV, oxygen is reduced at the platinum surface, initially to hydrogen peroxide H₂O₂ so that the polarity tends to discharge as electrons are donated to oxygen (which acts as an electron acceptor). The current which then flows is stoichiometrically related to the oxygen consumed at the cathode providing a fast, effective method of detecting small changes in oxygen tension in a liquid-phase sample.



DW2/2 electrode chamber

The DW2/2 electrode chamber has been developed specifically to allow simultaneous spectroscopic and oxygen flux measurements.

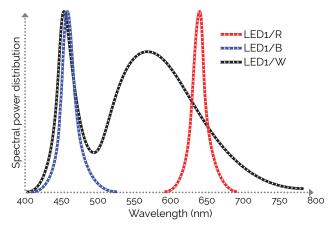
The reaction vessel of the DW2/2 is constructed from precision bore borosilicate glass tube with a prepared S1 electrode disc forming the floor of the vessel. Precision temperature control of sample and sensor is delivered via a concentric water jacket with suitable connection ports for a thermoregulated circulating water bath. The water jacket is constructed from black acetal which provides the ability for dark adaptation of samples or oxygen measurement in complete darkness. There are four optical ports mounted perpendicular to the reaction vessel at right angles to one another. The LED1 LED light source supplied with the Chlorolab 2+ system fits directly into any of these ports for uniform and high intensity illumination of the sample. Other light sources, fibre optic light guides and detectors may also be mounted on the DW2/2 via the ports enabling spectroscopic measurements to be made.

DW2/2 is fitted with a plunger with a capped central bore. The height of the plunger may be adjusted easily to suit liquid-phase sample volumes of between 0.2 - 2.5ml whilst the central bore easily accommodates Hamilton type syringes allowing additions/subtractions to/from the reaction vessel during an experiment.



LED1 light source

A LED1 high intensity light source in one of three colours is supplied with the Chlorolab 2+ system. The different light sources that can be supplied are as follows:



- LED1/R: red LED with a peak wavelength of 627nm
- LED1/B: blue LED with a peak wavelength of 470nm
- LED1/W: white LED with a colour temperature of 4100K

Light intensity adjustments are made automatically based on user-defined PFD tables within OxyTrace+ software. PFD tables consist of up to 20 individual steps allowing complex light response assays to be configured for automatic execution during a measurement.

Each of the LED1 light sources provide a maximum intensity of up to 2,000 μ mol m⁻² s⁻¹ in DW2/2. Greater intensities are achievable by purchasing additional LED1 housings and an adapter which allows simultaneous automatic control of up to 3 LED1 light units.

Quantitherm PAR/temperature sensor

The QRT1 consists of a handheld display unit combined with the QTP1+ probe sensor. For use as a calibration tool for the LED1 light source, the QTP1+ probe connects directly to the rear of the Oxylab+ control unit allowing a 10 step multi-intensity calibration routine within the OxyTrace+ software to be performed.

QRT1 may also be operated as a standalone sensor across the range of normal measuring temperatures used for both photosynthesis and cellular respiration studies and offers maximum accuracy in the 10°C – 40°C region. For photosynthesis measurements, the PAR quantum sensor provides a displayed resolution of 1µmol m⁻² s⁻¹ throughout the 0 to 5,000 µmol m⁻² s⁻¹ range and up to a maximum of 50,000 µmol m⁻² s⁻¹ with a displayed resolution of 10 µmol m⁻² s⁻¹.

OxyTrace+ software

OxyTrace+ is a multi-function Windows® program supplied with Chlorolab 2+ for system configuration, calibration, data acquisition and analysis. An automated 2 step calibration routine guides the user quickly and effectively through the system calibration process using electrode values measured from air-saturated and deoxygenated water.

OxyTrace+ allows simple configuration of comprehensive PFD tables consisting of up to 20 individual light steps. Light intensity adjustments are performed automatically during the measurement. OxyTrace+ also allows calibration of the LED1 light source from a simple software routine. This requires the QTP1 PAR/temperature sensor to be connected to the rear of the Oxylab+ control unit and placed into the reaction vessel prior to the addition of any liquids.

A tabbed interface allows a simple transition between the different data views including oxygen electrode (and if configured, auxiliary and external ion-selective electrode) real-time output, a split screen showing real-time rate of change above the oxygen signal and tabulated numerical data.

change above the oxygen signal and tabulated numerical data.

Post acquisition analysis tools allow automatic calculation of oxygen rates from user-defined rate intervals.

Additional analysis tools automatically calculate rates of change for defined PFD light steps with a calculation

data files opening effortlessly in external data processing packages such as MS Excel®.

of quantum yield presented at the end of a measurement. All files are saved as Comma Separated Values (CSV)

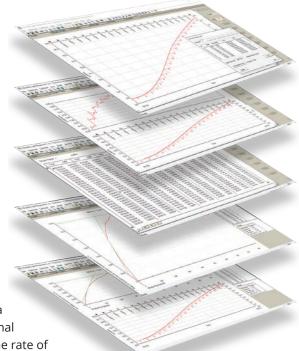
OxyTrace+ will run on all supported Microsoft operating systems.



Chlorolab 2+ systems are supplied with the following components:

- OXYL1+: Oxylab+ electrode control unit
- DW2/2: Oxygen electrode chamber
- S1: Oxygen electrode disc and SMB-SMB connection cable
- LED1: LED light housing in choice of 3 colours:
 - LED1/W: white LED (4100K default option)
 - LED1/R: red LED (627nm)
 - LED1/B: blue LED (470nm)
- QRT1: Quantitherm PAR/temperature sensor

- A2: Membrane applicator to assist with smooth application of electrode membrane
- S2/P: Pack of 5 magnetic followers
- S3: Pack of 2 replacement borosilicate glass reaction vessels
- S4: PTFE membrane (0.0125mm x 25mm x 33m)
- S6B: Set of replacement o-rings for DW2/2 electrode chamber
- S16: Cleaning kit for the S1 electrode disc.



Technical specifications

Oxylab+ electrode control unit

Measuring range: Oxygen: 0 - 100%, pH: 0 - 14pH,

Aux: 0 - 4.096V

Signal inputs: S1 O_2 electrode (SMB), pH/ISE (BNC),

Aux (8 pin Mini Din), QTP1 PAR/Temp

probe (6 pin Mini Din) Oxygen: 0.0003% (24 bit), pH: 0.0006pH (16 bit),

Aux: 62.5µV/bit (16 bit)

Polarising voltage: 700mV Input sensitivity: 0 - 9000nA

Resolution:

Magnetic stirrer: Software controlled 150 - 900rpm

in % steps

Sampling rate: 0.1 - 10 readings/s

Electronics: Microcontroller: 16 bit high

performance CPU running at 32 MHz

ADC: Dual, Low power, 16/24 Bit Sigma Delta

Display: 61 x 2 character blue LCD

Communications: USB2.0

Analogue output: $0 - 4.5 \text{V O}_2 \text{ signal}$

Dimensions (HWD): 250 x 125 x 65mm - 0.63 Kg
Power: 95 - 260V universal input mains supply. Output 12V DC 2.5A

DW2/2 electrode chamber

Suitability: Liquid-phase photosynthesis/

respiration

Construction: Black acetal

Sample chamber: Precision bore, borosilicate

glass tube

Sample volume: 0.2-2.5ml (1.5ml min. for illumination)
Plunger: Variable height, capped central bore

Temperature control: Water jacket connected to circulating

water bath

Optical ports: 4 x optical port with 16mm diameter

Dimensions: 105 x 65mm - 0.4Kg

S1 oxygen electrode disc

Electrode type: Clark type polarographic sensor Electrode output: Typically $1.6\mu A$ at $21\% O_2$ Residual current: Typically $0.04\mu A$ in $0\% O_2$ Response time: 10-90% typically <5 seconds Oxygen consumption: Typically $<0.015\mu mol/hr^{-1}$

LED1 light source

Cooling:

Light source: Single LED

Control: Via Oxylab+ & OxyTrace+ software Wavelength: LED1/W - 4100K (colour temperature)

LED1/R - 627nm peak wavelength LED1/B - 470nm peak wavelength Integral automatic cooling fan

Intensity: 2,000 μ mols m $^{-2}$ s $^{-1}$ (greater intensities

possible with multiple LED1 units)

Dimensions: 64 (dia) x 60 mm (h) - 270g

QRT1 PAR/Temperature sensor

Measuring range: $0-50000 \mu mol \ m^{-2} \ s^{-1} \ (+/-5\%) \ in \ 2$

ranges (0 - 5000 & 0 - 50000) in 400 -

700nm band

Resolution: 1μ mol m⁻² s⁻¹ at 0 - 5000

10μmol m⁻² s⁻¹ at 5001 - 50000

PAR sensor: Silicon photodiode & optical filter with

white acetal diffuser

Temperature sensor: RT curve matched bead thermistor.

0 - 50°C/32 - 122°F (0.02°C resolution) Handheld display unit. 16 x 2 LCD

Signal display: Handheld display unit. 16 x 2 LCD display. 0 - 5V analogue output of

PAR/ temperature values

Power requirements: 4 x 1.5V AA (LR6) cells

Typically 100 hours battery life

Dimensions (display): 146 x 92 x 32mm - 300g

Dimensions (QTP1): 9.5 x 107mm - 50g



Hansatech Instruments is a British company that has been developing high quality scientific instrumentation for over 40 years. Our systems are used widely for teaching & research in cellular respiration & photosynthesis programs in more than 100 countries throughout the world. We have gained an enviable reputation for quality, reliability & excellent price/performance.



Our product range consists of a range of modular solutions for the measurement of oxygen using Clark type polarographic sensors. We also develop chlorophyll fluorescence measurement systems using both continuous excitation & pulse-modulated measurement techniques with further optical instrumentation for the measurement of sample chlorophyll content.



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addition to instruments manuals
& software upgrades.

Hansatech Instruments Ltd

Narborough Road, Pentney, King's Lynn, Norfolk PE32 1JL, UK Tel: +44 (0)1760 338877 Fax: +44 (0)1760 337303

info@hansatech-instruments.com www.hansatech-instruments.com

